

# CURRENT

**Category:** General SOP for Extraction of genomic DNA from whole blood in Genetics laboratory.

**Title:** Extraction of genomic DNA from whole blood by phenol-chloroform method in the Department of Clinical Pharmacology, K.E.M Hospital, Mumbai.

**SOP No.:** Pre PCR-Method- 03

**Date first effective:** 1<sup>st</sup> January 2025

**Review date:** 31<sup>st</sup> December 2025

Department of Clinical Pharmacology, 1<sup>st</sup> floor, New MS building Seth GS Medical College & KEM Hospital, Parel, Mumbai 400012.

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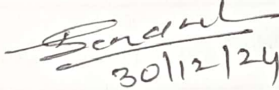
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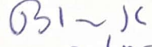
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**Author:** Dr. Sheetal Kudtarkar  
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**Signature with date:**   
30/12/24

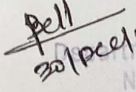
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30/DEC/2024

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**Signature with date:**

  
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**Purpose:** This standard operating procedure (SOP) describes the method for extraction of genomic DNA from whole blood.

**Scope:** This SOP is limited to extraction of genomic DNA from whole blood.

**Responsibilities:**

**Primary Responsibility:** Divya Bhare  
Lab Technician

*Bhare*  
30/Dec/2024

**Secondary Responsibility:** Dr. Sheetal Kudtarkar  
Project Scientist

*Sheetal*  
30/12/24

**References:** JIPMER (Pondicherry) Workshop 2012 manual



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### Detailed Instructions:

#### Important: This procedure should be carried out wearing sterile gloves

1. Collect 5mL blood in a 15mL fresh polypropylene tube containing 100µL of 10% EDTA disodium salt.
2. Collected blood sample is stored at -20°C and should be analyzed within two weeks.
3. Add 10mL of RBC lysis solution, mix vigorously for 5mins and keep the tube in deep freezer for 10mins (-20°C)
4. Centrifuge the tube at 2500 rpm in cooling centrifuge for 10 mins at 4°C
5. Discard the upper layer in bucket containing 10% hypochlorite solution by tilting the tube slowly in such a way that the pellet should remain in the tube
6. Add 6 to 8mL of RBC lysis solution, mix and follow the Steps 5,6 & 7 till the RBCs are lysed completely (**Till solution becomes clear; NOT more than 4 washes**)
7. Add 2.25 mL of WBC lysis solution in the tube. Mix gently with the help of rotospin machine for 20 mins at 21 rpm.
8. Add 125 µL of 10% Sodium dodecyl sulphate, 50 µL of Protienase K, and 75 µL of MilliQ water to it and mix it with the help of Rotospin for 20 mins at 21rpm.
9. Mix the above solution and incubate at 37°C for overnight in Incubator
10. Next day at 10:00am check whether the lysis is complete (If **lysis is complete the solution will be homogenous and will freely move when tilted to and fro**)
11. If lysis is incomplete invert the tube for 10 times gently and incubate at 37°C for 3- 4 hrs in Incubator
12. Add 1mL of saturated NaCl and mix.
13. Add 3.5mL of Chloroform and mix by to and fro movement (gently) with the help of Rotospin for 20 mins at 21 rpm till it becomes homogenous mixture.
14. Centrifuge at 4000 RPM at 4°C for 20 mins

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15. Take the upper layer by micropipette (**by slightly rotating the pipette by the sides of the wall of the test tube**) without disturbing the lower layer and transfer the upper layer to a fresh tube (**use new tips for each sample**)
16. Add 2.5mL of equilibrated phenol. Mix gently with the help of rotospin for 20 mins at 21 rpm and centrifuge at 4000 RPM for 20 min at 4°C
17. Take upper layer using micro pipette without disturbing lower layer and transfer to a fresh tube.
18. Add 3 mL of Chloroform: Octanol (**ratio 24:1**) and mix gently with the help of Rotospin for 20 mins at 21 rpm and centrifuge at 4000 RPM for 20 min at 4°C
19. Transfer the upper layer to a fresh tube (**If the interphase is not clear at this stage keep the tube standing for 5-10 mins**) Interphase: A thin layer between upper and lower layers
20. Add 6mL of ice-cold absolute ethanol which is kept in ice box in -20°C deep freezer and mix by inverting the tubes for 6 to 8 times. High molecular weight DNA will precipitate at this step
21. Remove DNA using a clean pipette tip and wash it with 70% ethanol in 1.5 mL v-shaped polypropylene tube by tapping the tube five to six times
22. Remove the DNA from the ethanol tube with a pipette and air dry
23. Transfer the DNA in a 1.5mL v-shaped micro centrifuge tube containing 200 µL of Tris EDTA buffer (pH 7.4).
24. Incubate at 37°C for 2-3 days in Incubator
25. Prepare two aliquots
26. Store the one aliquot of DNA sample at -20°C (**for 4 to 5 years**) and the second aliquot at 4°C (**one year**)



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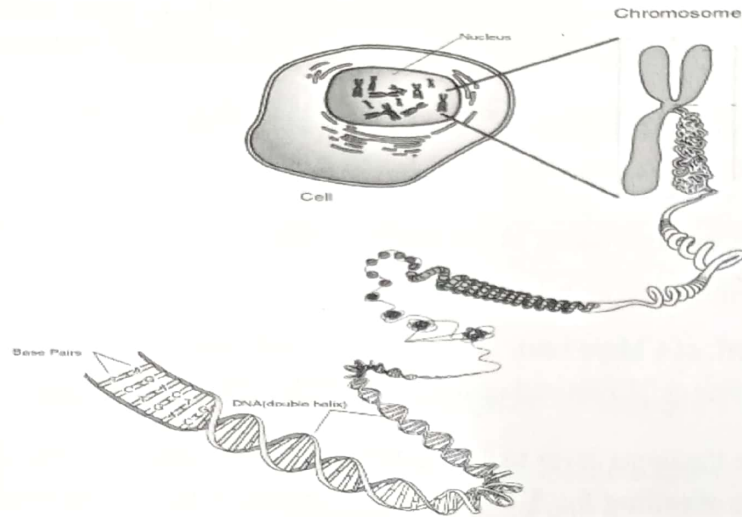
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**DNA Structure:**



**SUGAR-PHOSPHATE BACKBONE OF DNA**

**FOUR BASES AS BASE PAIRS OF DNA**

**ELECTRON MICROGRAPH OF DNA**

**DNA DOUBLE HELIX**

1 helical turn = 3.4 nm

minor groove major groove